

A new Eriophyoid mite causing damage to creeping fig plant, from India

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Abstract

Eriophyoid mites are tiny microscopic arachnids of the order Acari. They are obligatory plant parasites and cause damage to the plant by their sap sucking habits. During routine taxonomic field work particularly for this group of mites in the sub Himalayan plains of Northeast India, a new parasitic mite *Diptilomiopus ficicolus* n. sp. belonging to family Diptilomiopidae is noticed along the ventral surface of the leaves of *Ficus heterophylla* L. (Moraceae). *Ficus heterophylla*, also known as Creeping fig, holds importance due to its medicinal uses, with parts of the plant used to treat conditions like rheumatism, ear infections, and as a poultice. The new species of the mite produces damage to the leaves of this beneficial plant by its sucking habit. This new mite species is studied thoroughly from the taxonomic point of view and described with proper illustrations from the collected specimen from northern district of West Bengal, India (Latitude: 24°50'40" N and Longitude: 87°55'50" E). After proper taxonomic diagnosis, the new mite is named according to the rules laid by the ICZN and the type specimens are registered in ZooBank account. The relationship of mite with its host plant is also discussed in this article. Taxonomic knowledge of the parasitic mite is very much essential for the proper eradication and control of this pest species.

Keywords: Plant mite, *Diptilomiopus ficicolus*, new species, *Ficus heterophylla*, Taxonomy, India.

Introduction

The super family Eriophyoidea of order Acari of class Arachnida includes ticks and mites. Mites belonging to the super family Eriophyoidea are entirely phytophagous and form a microenvironment in various plant parts where they live. (Shukla, 2021^[32], Vervaeke, 2021^[34], Jeppson *et al.*, 1975). They are morphologically as well as biologically highly specialized parasites that wonder biologists around the world. They are highly host specific and they show an intimate host parasite relationships (Abdel-Khalek & Momen, 2022^[1]; Brown *et al.*, 2021, Oldfield, 2010)^[26]. During their feeding on plant sap and they produce mechanical injury to the plant parts and are known to transmit various plant viruses along with their saliva they pour into it. Various damage symptoms like gall formation, hairy outgrowths, witches broom, yellowing of leaves, brown patchy are most common. (Druciarek *et al.*, 2019; Sarwar, 2020^[28]; Stephan *et al.*, 2008)^[31]. Keifer *et al.* (routine survey is conducted for these mites in northern parts of west Bengal, India in the months of September to November, 2024. The detailed taxonomic study is done on the collection made on *Ficus heterophylla* from this locality a new mite species is found in the genus *Diptilomiopus*. This genus was erected by Nalepa in the year 1916. (see Nalepa 1916)^[25]. *Ficus heterophylla*, known as the Creeping fig, is a tropical tree native to Southeast Asia and the Pacific Islands, with distinctive leaves that vary in shape and size, ranging from lobed to entire forms (Fig 1A.) It is also cultivated as an ornamental tree and has medicinal uses, with its bark and root powder used for cough, asthma, and chest pain (Ghugre 2024, Hamilton 2004, Devi *et al.*, 2022, Dhanya *et al.*, 2012 Sharma *et al.*, 2014^[30], Kumar *et al.* 2002^[23], Satapathy and Kumar 2017)^[29]. Fruits are eaten as vegetable item by the local communities. The new species causes severe damage to the tender leaves of the plant. As of

2024, according to working Catalogue of Eriophyoidea of the World, Versio:10, 'The catalogue of the Eriophyidae' (Joel Hallan; biocat@ccms.net), the genus *Diptilomiopus* includes 104 valid species within it. (Sur *et al.*, 2018^[33]; Craemer *et al.*, 2017; Amrine and Stasny 1994, Amrine *et al.*, 2003, Yan-mei yuan- Xiao-Feng Xue 2019^[35], Chakrabarti *et al.*, 2019). As of now, plant family Moraceae harbors 9 species and Anacardiaceae harbors 5 species of this genus *Diptilomiopus* in India (Chakrabarti *et al.*, 2019). From India one new species is added to this genus from this taxonomic research. As per taxonomic procedure the new mite species in this genus is named following ICZN rules and registration of the new species is done in Zoo-Bank Account. The holotype specimen has been kept in laboratory of Entomology Research Unit of the Serampore College of Calcutta University, a national repository recognized by ZSI.

Materials and Methods

The mite infested leaves are examined under stereomicroscope. Then the permanent slides of these specimens are prepared following the standard taxonomic procedure as stated by Amrine *et al.* 2003^[4] and subsequently modified by Lillo *et al.*, 2010. Mites are mounted in Hoyer's medium (Jeppson *et al.*, 1975). After preparation of slides the specimens are studied in phase contrast microscope Model: Letiz Dialux 20. A camera Lucida line drawings of whole mite and different parts of the mites having taxonomic importance are made (Fig.1A-G). Measurements (as stated by Amrine and Manson, 1996)^[3] of different body parts and the length of the setae are taken in micrometer (μm). In the text the morphological terminology and abbreviations are used as per Lindquist *et al.*, (1996)^[24]. In the text the measurements of the different morphological and anatomical characters of the holotype

specimens are flowed by the ranges of measurements of paratype specimens given in the brackets. All the slides bearing holotype and paratypes are now kept in Entomology laboratory of Serampore College of University of Calcutta. According to the ICZN Rules, after the publication of the new species the type specimen will be deposited in National Collections of Zoological Survey of India, Head Quarter in Kolkata, a National repository.

Results and Discussions

Taxonomic studies and differential diagnosis with the preexisting species of this genus reveals this mite species new to Science. The etymology of the new name of the species is given after the ICZN Rules and the Registration of new species in Zoo-Bank is properly done. The account link is generated here: urn:lsid:zoobank.org:pub:FF1E2FCE-A7FE-46C2-BB0B-E7F9D558AA0D.

Description of the Genus: *Diptilomiopus Nalepa 1916* ^[25]

Body is robust, spindle shaped. Gnathosoma and chelicerae large and set at perpendicular or strongly recurved in between the leg I. Prodorsal shield wider than long, without frontal lobe, scapular tubercles and setae usually absent, in some species scapular tubercles without seta may be present. Legs with distinct genu or fused with femur, genual seta absent; only tarsal setae present on legs; tarsal empodium deeply divided. Coxae I may or may not be separated, with less prominent sternal line; *1b* tubercles absent; opisthosoma sub circular in cross section with 1 to 3 ridges just behind the prodorsal shield; seta *c2* missing, often tubercles present; epigynium may be smooth or provided with longitudinal ribs or lines and granules or tubercles; internal apodeme normal in length.

Type species: *Diptilomiopus javanicus* Nalepa, 1916 ^[25].

Taxonomic description of new species: *Diptilomiopus ficicolus* sp. nov. (Figs2. A-G)

[Zoo-Bank:urn:lsid:zoobank.org:pub:FF1E2FCE-A7FE-46C2-BB0B-E7F9D558AA0D]

Female (1 holotype 67 paratype) (Figs. 2): Reddish in colour, 123.2 (123.2-124.1) long, 62.5 (62.5-61.6) wide, robust, fusiform with 4 ridges running upto two-third of the body length. *Gnathosoma* 16.8 (15.8-16.8) long and curved downward, dorsal pedipalp genual seta is 3.7 (3.3-3.7) long.

Prodorsal shield (Fig 2B: Antero-dorsal mite) 23.3 (22.8-23.3) long and 50.4 (50.4-51.3) in width, prodorsal shield design (Fig. 1B) shows a network of 22 closed cells including a central large polygonal cell, number and size of cells show bilateral symmetry, median line incomplete as the central cell do not have median line, admedian line complete and criss-crossing the entire length of prodorsal shield, sub-median lines also sinuate, each half of the prodorsal shield shows 5 anterolateral cells; scapular tubercle and seta *sc* absent.

Opisthosoma (Fig 2.A; lateral Mite) with 52 (52-54) long and dorsal annuli are smooth, 69 (69-72) ventral annuli bear micro tubercles. Rounded microtubercles are located on anterior margins of ventral annuli; last 8 ventral annuli with micro striations; seta *c2* absent, seta *d* 10.2 (10.2-10.7) long on annulus 26 (26-27); seta *e* 7.4 (6.5-7.4) long and located

on annulus 41; Seta *f* is 35.4 (34.5-35.4) in length and is located on 61st ventral annulus; seta *h1* is not present and seta *h2* is 33.60 (33.6-34.5) in length.

Leg I (Fig. 2C) from base of trochanter 36.4 (36.4-37.3) long; femur 14.0 (13.0-14.0) long, without basiventral femoral seta *bv*; genu fused with femur. Tibia is 5.6 (5.1-5.6) long and paraxial tibial seta *l'* absent; 3.7 (3.7-4.2) long tarsus with two identical paraxial fastigial tarsal seta *ft'* and antaxial fastigial tarsal seta *ft''* 22.4 (22.4-23.3). paraxial unguinal tarsal seta *u'* is 4.6 (3.5- 4.7), slightly curved tarsal solenidion ω is knobbed and 5.6 (4.5- 5.7) ; tarsal empodium *em* is divided and with 6, tarsal empodium *em* is 7.3 (7.1-7.4).

Leg II (Fig 2.D) near the base of trochanter it is 3.4.5 (34.5-35.4); genu is completely fused with femur and 14.0 (13.8-14.0) ; basiventral femoral setae *bv* is absent; 2.8 (2.8-3.7) long tibia without paraxial tibial seta *l'*; 5.6 (5.1-5.6) long tarsus with, paraxial fastigial tarsal seta *ft'* 18.6 (17.7-18.6), antaxial fastigial tarsal seta *ft''* is lacking; the length of paraxial unguinal tarsal seta is 4.6 (3.7-4.6); tarsal solenidion ω 4.6 (4.6-5.5) long;

Tarsal empodium (Fig. 2E.): 6 rayed divided tarsal empodium *em* 7.4 (6.5-7.4) long.

Coxal-genital region (Fig.2F.): Coxae I 14.9 (14.9-15.4) long and contiguous basally; coxal surface smooth; *1b* tubercles with seta absent but *1a* tubercles with seta present a little ahead of the line across the *2a* tubercles with seta; 12.1 (12.1-12.5) long seta *1a*. Coxa II 13.0 (13.0-14.0); Seta *2a* is 23.4 (23.3-24.1). Genitalia 14.0 (14.0-14.9), 15.8 (15.8-16.3) wide; epigynium ellipsoidal in shape, with anterolateral scorings, a median Y shaped scoring is seen, seta *3a* 4.6 (4.6-5.5) long.

Internal genitalia (Fig.1.G): It shows two round spermatheca and branched oviduct

Male: Males are not observed as the important taxonomic variables of this mite are found only in case of female individuals.

Type material: Holotype: Female (marked) on slide (no. 1418/76/2024), India: West Bengal: Malda, English bazar, Latitude: 24°50'40" N and Longitude: 87°55'50" E, 14.x.2024 from *Ficus heterophylla* L.(Moraceae), Coll. S. Sarkar. Paratypes: 13 females on slide bearing holotype and 54 females on 6 slides (nos. 1419-1424/76/2024), collection data same as in holotype.

Type Locality: India: West Bengal: Malda, English bazar, Latitude: 24°50'40" N and Longitude: 87°55'50" E

Relation to the host plant: Mites are found in maximum numbers on the ventral surface of leaves near the mid vein. Due to its sap sucking habit yellowing of leaves occurs and in severe infestation brown spots are seen on leaves. Shoots are eventually dried up. (Fig. 1C)

Etymology: The specific epithet of the name "*ficicolus*" is derived from combination of two different words, one is '*Ficus*'- the generic name of the host plant and other is a Latin word '*Colus*' means inhabitant or dwelling in. Thus the meaning of specific epithet '*ficicolus*' is the inhabitant of *Ficus* plant. From this binomial nomenclature one can easily understand the host of this mite.

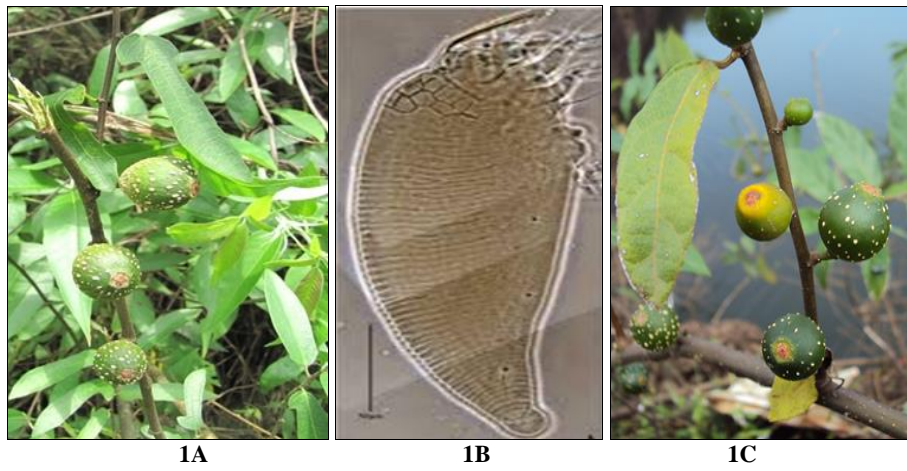


Fig: 1A. Fresh shoot of *Ficus heterophylla*; **1.B.** Photograph of the mite taken through phase contrast microscope; **1C.** Mite infested damaged shoot of *Ficus heterophylla*

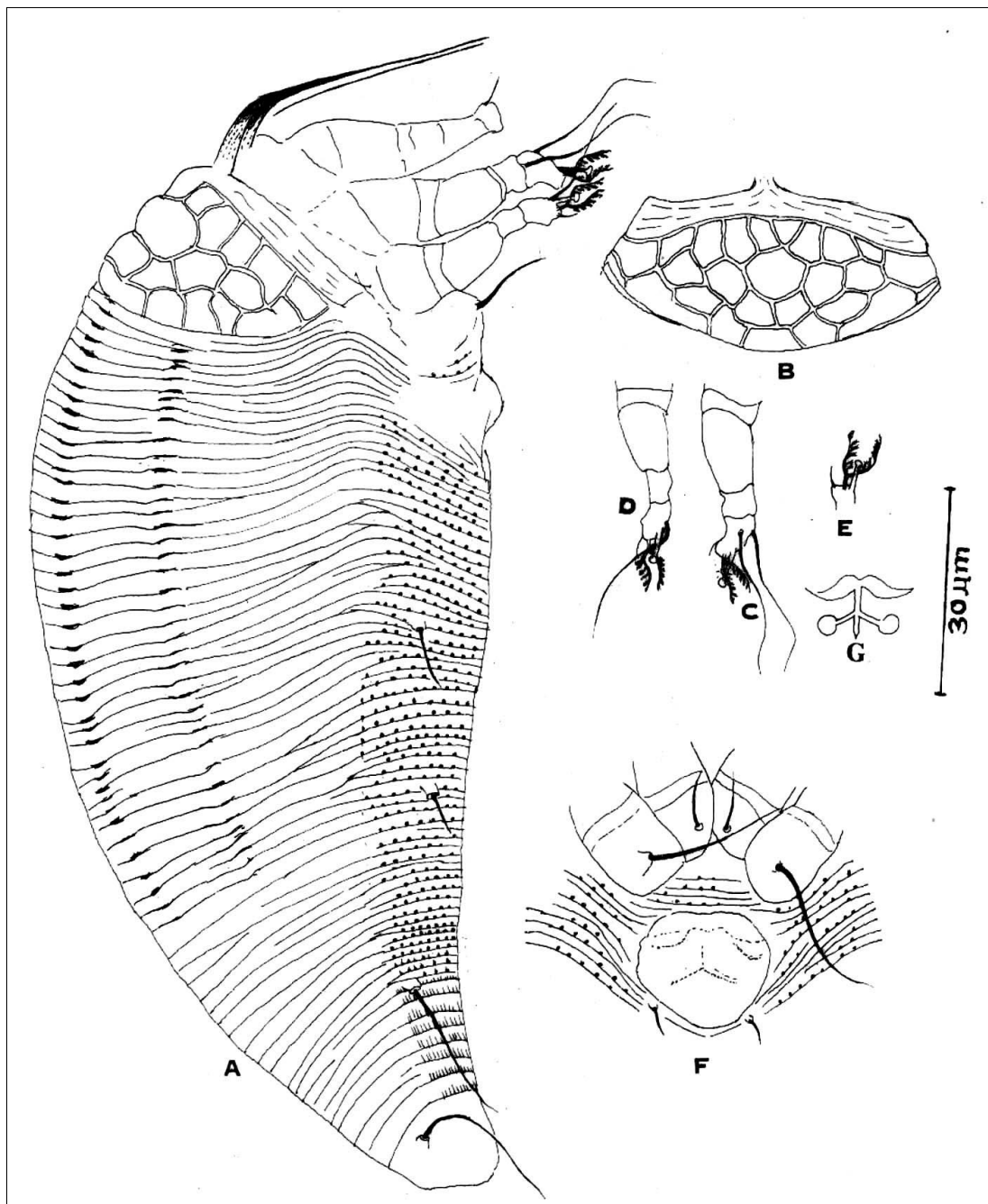


Fig 2: *Diptilomiopus ficicolus* sp.nov, holotype, no. 1418/76/2024: A-Antero-dorsal mite (AD); B- Antero-Dorsal mite (AD); C- Leg I (L1); D- Leg II (L2); E- Tarsal empodium of L1 (em); F- Coxal-genital region (CG); G- Internal genitalia of female.

Taxonomic diagnosis

The species in *Diptilomiopus* fall under two distinguished group, first group having central cell with radiating lines on prodorsal shield and the second group having network of cells on prodorsal shield. The new species falls in the second group and shows similarities with *D. davisii* Keifer (1969), *D. holmesi* Keifer (1962), *D. javeremovici* Keifer (1960) and other 12 Indian species but differs from them by the numbers of closed cells on the dorsal shield. Among Indian species, this new species comes very close to *D. trewir* Chakrabarti and Mondal (1983) and *D. ficusis* Chakrabarti and Mondal (1983) in absence of scapular tubercles and scapular setae, but differs from them by the presence of 6 rayed tarsal empodium.

Conclusion

The new mite species discovered from this area appears a potential pest of *Ficus heterphylla* an important medicinal as well as vegetable yielding plant in India. This mite pest produces damage to the leaves by its feeding habit and parasitic mode of living. This finding may be helpful to provide the basis to the further taxonomic study including DNA Barcoding. Entomologist of plant protection department may identify the mite pest and the nature of damage through this research outcome.

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References

1. Abdel-Khalek AA, Momen FM. Biology and life table parameters of *Proprioseiopsis lindquisti* on three eriophyid mites (Acari: Phytoseiidae: Eriophyidae). *Persian Journal of Acarology*,2022;11(1):59–69. <https://doi.org/10.22073/pja.v11i1.68574>
2. Amrine JW Jr, Stasny TA. *Catalog of the Eriophyoidea (Acarina: Prostigmata) of the world*. Michigan: Indira Publishing Houses, 1994, 798.
3. Amrine JW Jr, Manson DCM. Preparation, mounting and descriptive study of Eriophyoid mites. In: Lindquist EE, Sabelis MW, Bruin J, editors. *Eriophyoid mites. Their Biology, Natural Enemies and Control*. World Crop Pests, Amsterdam: Elsevier Science Publishers,1996;6:383–396. doi:10.1016/S1572-4379(96)80023-6
4. Amrine JW Jr, Stasny TA, Flechtmann CHW. Revised keys to world genera of Eriophyoidea (Acari: Prostigmata). Michigan: Indira Publishing Houses, 2003, 244.
5. Brown MS, Blubaugh CK, Chong JH. Biology and management of eriophyid mites in turfgrass. *Journal of Integrated Pest Management*,2021;12(1):25. <http://dx.doi.org/10.1093/jipm/pmab020>
6. Chakrabarti S, Mondal S. An Account of the Genus *Diptilomiopus* Nalepa (Acarina: Eriophyoidea) from India with Descriptions of three new species and key to Indian species. *Acarologia*,1983;24(3):299–308.
7. Chakrabarti S, Sur S, Roy S, Sarkar S. Two new genera and two new species of Eriophyoid mites (Acari: Eriophyoidea) from North Bengal, India. *Zootaxa*,2017;4236(1):172–182. doi:10.11646/zootaxa.4236.1.10
8. Chakrabarti S, Sur S, Sarkar S. Two new species of *Diptilomiopus* Nalepa (Acari: Eriophyoidea) from India. *Acarologia*,2019;59(3):383–394. <https://doi.org/10.24349/acarologia/20194337>
9. Craemer C, Amrine JW Jr, Childers CC, Rogers ME, Achor DS. A new eriophyoid mite species, *Diptilomiopus floridanus* (Acari: Eriophyoidea: Diptilomiopidae), from citrus in Florida, USA. *Systematic & Applied Acarology*,2017;22(3):386–402. doi:10.11158/saa.22.3.5
10. de Lillo E, Craemer C, Amrine JW Jr, Nuzzaci G. Recommended procedures and techniques for morphological studies of Eriophyoidea (Acari: Prostigmata). *Experimental & Applied Acarology*,2010;51:283–307. doi:10.1007/s10493-009-9311-x
11. Devi R, Manjula BL, Kumar M, Kumar S, Marndi S. Food and medicinal values of some *Ficus* species in Medico-Biowealth of India, Volume-VI,2022. ISBN:978-81-952750-9-0. doi:10.5281/zenodo.6877024
12. Dhanya B, Viswanath S, Purushitham S. *Ficus* trees in rainfed agricultural systems of Karnataka, southern India: An analysis of structure, benefits, and farmers' perceptions,2012;50(1-2):59–62.
13. Druciarek T, Lewandowski M, Tzanetakis I. A new, sensitive and efficient method for taxonomic placement in the Eriophyoidea and virus detection in individual eriophyoids. *Experimental and Applied Acarology*,2019;78(2):247–261.
14. Ghuge N, Shegar D, Jadhav S, Nagare V. *International Journal of Pharmaceutical Sciences*,2024;2(12). doi.org/10.5281/zenodo.1441137. Article Id IJPS/240212052
15. Hamilton AC. Medicinal plants, Conservation, and Livelihoods. *Biodiversity and Conservation*,2004;13:1477–1517.
16. Jeppson LR, Keifer HH, Baker EW. *Mites Injurious to Economic Plants*. Berkeley: University of California Press, 1975, 614.
17. Joel Hallan (biocat@ccms.net). *A Working Catalogue of the Eriophyoidea of the World*. Version 1.0. The Catalogue of the Eriophyidae. Accessed on 05 March 2025.
18. Keifer HH. Eriophyid studies B-1. *Bur Entomol, Calif Dept Agr*, 1960, 20.
19. Keifer HH. Eriophyid studies B-7. *Bur Entomol, Calif Dept Agr*,1962, 20.
20. Keifer HH. Eriophyid studies C-1. *ARS-USDA*, 1969, 24.
21. Keifer HH, Baker EW, Kono T, Delfinado M, Styer WE. An illustrated guide to plant abnormalities caused by Eriophyid mites in North America. *USDA Agricultural Handbook No. 573*,1982:178 p.
22. Kumar M, Sharma I, Verma PK, Singh BJ, Singh R, Upadhyay SK. A study on diversity and distribution of *Ficus* L. (Dicotyledonae: Moraceae) species at Forest Research Institute (FRI), Dehradun (Uttarakhand), India. *Journal of Applied and Natural Science*,2021;13(2):552–560.

23. Kumar SN, Mishra S, Kumar S. A note on *Ficus hederacea* Roxb. (Moraceae). *Species*,2022;23(71):144–147.
24. Lindquist EE. External anatomy and notation of structures. In: Lindquist EE, Sabelis MW, Bruin J, editors. *Eriophyoid mites: their biology, natural enemies and control*. World Crop Pests, vol. 6. Amsterdam: Elsevier Science Publishers, 1996, 3–31.
25. Nalepa A. Neue Gallmilben (32. Fortsetzung). *Anzeiger der kaiserlichen Akademie der Wissenschaften, Mathematisch-naturwissenschaftliche Klasse, Wein*,1916;53(22):283–284.
26. Oldfield G, Skoracka A, Smith L, Cristofaro M, Amrine JW Jr. Host-plant specificity and specialization in eriophyoid mites and their importance for the use of eriophyoid mites as biocontrol agents of weeds. *Experimental and Applied Acarology*,2010;51:93–113. doi:10.1007/s10493-009-9323-6
27. Sarkar S. Eriophyoid mites (Acari) of Malda and Dakhin Dinajpur of West Bengal, India [PhD Thesis]. Kalyani: University of Kalyani, 2011,160 p.
28. Sarwar M. Mite (Acari: Acarina) vectors involved in transmission of plant viruses. In: *Applied Plant Virology*. Elsevier, 2020. p,257–273. <http://dx.doi.org/10.1016/B978-0-12-818654-1.00020-7>
29. Satapathy S, Kumar S. Medicinal *Ficus* species of temple city of Odisha. *Journal of Biodiversity and Conservation*,2017;1(3):1–5.
30. Sharma M, Sharma CL, Lalmalsawma M, Singh MK, Gogoi BR. Wood anatomy of some *Ficus* species of Mizoram, NE India with reference to their identification. *International Journal of Botany and Research*,2014;4(2):19–30.
31. Stephan D, Moeller I, Skoracka A, Ehrig F, Maiss E. Eriophyid mite transmission and host range of a *Brome streak mosaic virus* isolate derived from a full-length cDNA clone. *Archives of Virology*,2008;153:181–185. <https://doi.org/10.1007/s00705-007-1065-3>
32. Shukla A. Mites. In: *Polyphagous Pests of Crops*, 2021, 409–455.
33. Sur S, Roy S, Chakrabarti S. Two new eriophyoid mites (Acari: Eriophyoidea) from West Bengal, India. *Zootaxa*,2018;4434:193–200. doi:10.11646/zootaxa.4434.1.13
34. Vervae L, De Vis R, De Clercq P, Van Leeuwen T. Is the emerging mite pest *Aculops lycopersici* controllable? Global and genome-based insights in its biology and management. *Pest Management Science*,2021;77(6):2635–2644. <https://doi.org/10.1002/ps.6265>
35. Yuan Y, Xue XF. Two new species of eriophyid mites (Acari: Eriophyidae) from Malaysia. *Zootaxa*,2019;4613(1):152. doi:10.11646/zootaxa.4613.1.8