

## Community composition of aquatic insects in relation to water quality in a pond in Midnapore town, West Bengal, India

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### Abstract

Twenty five species of aquatic insects have been recorded from a small weed infested water body in Midnapore town, West Bengal, India. Hemiptera was the most abundant order comprising 51.93% of total entomofauna followed by Odonata (16.12%), Coleoptera (15.15%), Ephemeroptera (13.68%) and Diptera (3.12%) in that order. *Anisops breddini* was the only dominant species along with 11 subdominant, 12 recedent and one sub-recedent species. Most of the species were found to be either positively or negatively correlated with the physico-chemical properties of the water. Existence of 13 species of macrophytes provided ample scope of niche differentiation for the coexistence of the insect species.

**Keywords:** Entomofauna, dominance status, correlation with water parameters

### Introduction

Aquatic insects play important role not only in the trophic dynamics of the ecosystem but also in the indication of changes in the quality of habitats and their degradation because of their ability to respond quickly to such changes and thus serve as reliable bioindicator of aquatic ecosystem [1]. In India about 5000 species of insects are estimated to inhabit inland waterbodies [2]. Most of these constitute the food for many commercially important fish while some are predacious, feeding upon the spawn and fry or competing with them for natural food. Several species are known to predate upon mosquito larvae and are used as potential biocontrolling agent for mosquito larvae [3, 4]. Aquatic insects also play a crucial role in providing various ecosystem services [5]. Some notable contributions on aquatic insects in recent years are those by Sharma and Agarwal [6], Majumder *et al.* [7], Barman and Baruah [8], Choudhury and Gupta [9], Choudhary and Ahi [10]. Work done in undivided Midnapore district and West Midnapore districts are those by Pahari *et al.* [11-15] and Jana *et al.* [16]. In

the present study an investigation has been undertaken to analyse the insect diversity and community structure of a weed infested man made pond.

### Materials and Methods

The present investigation was carried out in a manmade perennial pond (22°25'08.7"N 87°19'54.2"E), 1134.75 m<sup>2</sup> with an average depth of 5.4 meter (Figure 1). The water body was infested with 13 species of aquatic weeds *viz.*, *Eclipta alba* Hassk., *Nymphaea nouchali* Burm., *Marsilea minuta* Linn., *Nymphoides indica* (Linn.), *Eichhornia crassipes* (Mart.), *Commelina benghalensis* Linn., *Hydrilla verticillate* Casp., *Vallisneria spiralis* Linn., *Ceratophyllum demersum* L., *Ipomoea carnea* (Mart.), *I. aquatica* Forrsk., *Hygrophila schulli* (Hamilt.), *Lemna perpusilla* (Torr.). Insects were collected at monthly interval from January 2020 to December 2020 between 7 am to 10 am from four sampling sites of the pond using a circular net with an area of 4208 cm<sup>2</sup> having 245 µm mesh size. Collected samples were preserved in 70% ethanol.

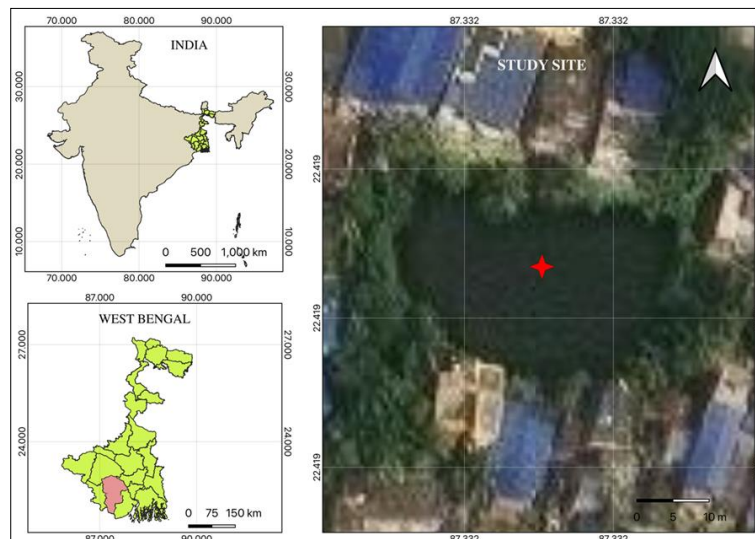


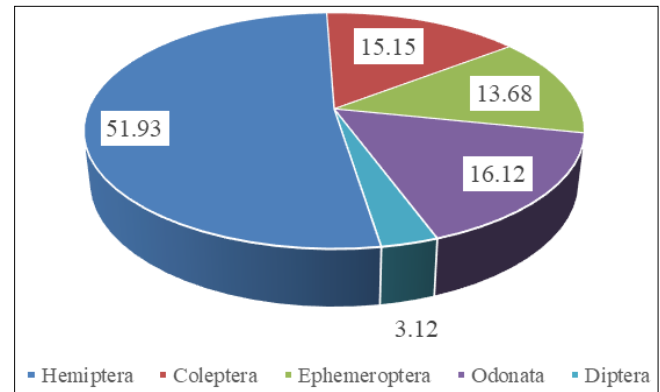
Fig 1: Study site

Aquatic insects were identified up to species level. Dominance status of the each species was determined through the analysis of relative abundance following Englemann [17]. Dissolve oxygen (DO), free carbon dioxide (CO<sub>2</sub>), Biological oxygen demand (BOD) and total alkalinity (TA) were measured following APHA [18]. Temperature and pH were measured by portable digital meters. Statistical analyses were done by SPSS (Ver. 26), and Ms Excel 2021 software.

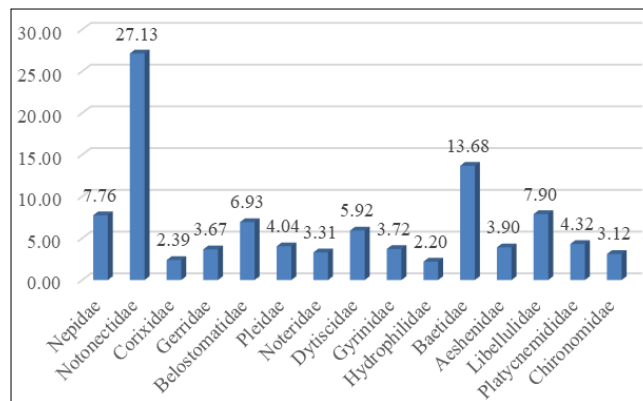
**Result and Discussion**

25 species under 5 orders, 15 families and 20 genera were recorded from the study site. Hemiptera was the most common order comprising 52.93% of the total recorded aquatic insects. This order was represented by 6 families and 10 species. Preponderance of Hemiptera in aquatic ecosystem has also been reported by Majumdar *et al.* [7] in Tripura, Barman and Baruah [8] in Assam Choudhury and Gupta [9] and Pahari *et al.* [13] in Purba Medinipur district, West Bengal. Diversity and abundance of macrophytes is known to influence the abundance and diversity of aquatic hemipteran insect Pahari *et al.* [14]. Odonata was the second most abundant order (16.12%) represented by 3 families and 6 species. This was followed by Coleoptera (15.15%), Ephemeroptera (13.68%), Diptera (3.12%) in that order (Figure 2). Order Coleoptera was represented by 4 families and 6 species while Ephemeroptera and Diptera were represented by a single family each and 2 and 1 species respectively. Family level analysis revealed that Family Notonectidae (Order Hemiptera) was the most

predominating family (27.13%) followed by Baetidae (13.68%), Libellulidae (7.90%), Nepidae (7.76%), Belostomatidae (6.93%), Dytiscidae (5.92%), Platycnemididae (4.32%), Pleidae (4.04%), Aeshenidae (3.90%), Gyrinidae (3.72%), Chironomidae (3.12%), Corixidae (2.39%) and Hydrophilidae (2.20%) in that order respectively (Figure 3). Order Odonata is also known to prefer freshwater habitats with luxuriant growth of macrophytes [10]. Turning to species wise analysis it was seen that *A. breddini* was the only dominant species which might be considered as the best exploiter in that pond. There were also 11 subdominant, 12 recedent and 1 sub-recedent species in that pond (Table 1).



**Fig 2:** Relative abundance of various insect Orders (%)



**Fig 3:** Number and relative abundance (%) of Families

**Table 1:** Abundance, relative abundance (%) and dominance status of species

Sl. No.	Species	X±SE	RA	DS
Order: Hemiptera				
Family: Nepidae				
1	<i>Laccotrephes ruber</i> (Linnaeus, 1764)	2.29±0.35	2.53	R
2	<i>Ranatra filiformis</i> Fabricius, 1790	3±0.32	3.31	SD
3	<i>Ranatra sordidula</i> Dohra, 1860	1.75±0.26	1.93	R
Family: Notonectidae				
4	<i>Anisops breddini</i> Kirkaldy, 1901	24.63±3.04	27.13	D
Family: Corixidae				
5	<i>Micronecta scutellaris</i> (Stål, 1858)	2.17±0.41	2.39	R
Family: Gerridae				
6	<i>Gerris spinolae</i> Lethierry & Severin 1896	3.33±0.42	3.67	SD
Family: Belostomatidae				
7	<i>Diplonychus annulatum</i> (Fabricius, 1781)	3.54±0.44	3.90	SD
8	<i>Diplonychus rusticus</i> (Fabricius, 1781)	2.08±0.37	2.30	R
9	<i>Lethocerus indicus</i> (Lepelletier & Serville, 1825)	0.67±0.12	0.73	SR
Family: Pleidae				

10	<i>Plea liturata</i> (Fieber, 1844)	3.66±0.46	4.04	SD
Order: Coleoptera				
Family: Noteridae				
11	<i>Canthydrus luctuosus</i> (Aubé, 1838)	3±0.33	3.31	SD
Family: Dytiscidae				
12	<i>Laccophilus purvulus</i> Aubé, 1838	3.17±0.46	3.49	SD
13	<i>Hydrovatus bonvouloiri</i> Sharp, 1882	2.21±0.28	2.43	R
Family: Gyrinidae				
14	<i>Orectochilus cylindricus</i> Régimbart, 1892	1.75±0.43	1.93	R
15	<i>Orectochilus cardoni</i> Régimbart, 1892	1.63±0.38	1.79	R
Family: Hydrophilidae				
16	<i>Sternolophus rufipes</i> (Fabricius, 1792)	2±0.39	2.20	R
Order: Ephemeroptera				
Family: Baetidae				
17	<i>Cloeon bicolor</i> Kimmins, 1947	7.83±1.91	8.63	SD
18	<i>Cloeon bengalense</i> Kimmins, 1947	4.58±1.14	5.05	SD
Order: Odonata				
Family: Aeshenidae				
19	<i>Anaciaeschna jaspidea</i> (Burmeister, 1839)	1.75±0.24	1.93	R
20	<i>Anax guttatus</i> (Burmeister, 1839)	1.79±0.28	1.97	R
Family: Libellulidae				
21	<i>Brachythemis contaminata</i> (Fabricius, 1793)	3.29±0.41	3.63	SD
22	<i>Crocothemis servilia</i> (Drury, 1773)	2.25±0.24	2.48	R
23	<i>Pantala flavescens</i> (Fabricius, 1798)	1.63±0.18	1.79	R
Family: Platynemididae				
24	<i>Copera marginipes</i> (Rambur, 1842)	3.92±0.36	4.32	SD
Order: Diptera				
Family: Chironomidae				
25	<i>Chironomus plumosus</i> (Linnaeus, 1758)	2.83±0.55	3.12	SD

[Relative abundance (RA) <1 = Subprecedent (SR); 1–3.1 = Recedent (R); 3.2–10 = Subdominant (SD); 10.1–31.6 = Dominant (D); >31.7 = Eudominant (ED)]<sup>[10]</sup>.

Aquatic insects are good indicator of water quality and they are mostly affected by the nature of the waterbody<sup>[10]</sup>. Analysis of the simple correlation revealed that most of the species viz., *L. ruber*, *R. sordidula*, *A. breddini*, *M. scutellaris*, *D. rusticus*, *C. luctuosus*, *L. purvulus*, *O. cyndricus*, *C. servilia* were positively correlated with water temperature. 2 ephemeropteran species viz., *C. bicolor* and *C. bengalense* on the contrary, were found to be negatively correlated with the water temperature. Water temperature is an important physical factor that influence growth rate<sup>[19-22]</sup> and success of egg development which ultimately leads to higher abundance of aquatic insects in higher temperature<sup>[23-25]</sup>. Significant negative correlation of Ephemeroptera with water temperature as in present study has also been previously shown by Choudhary and Ahi<sup>[10]</sup> in Madhya Pradesh. pH plays an important role in many biological and chemical properties of freshwater<sup>[26]</sup>. Many species in the present study viz., *L. ruber*, *R. sordidula*, *M. scutellaris*, *D. annulatum*, *L. indicus*, *P. liturata*, *C. luctuosus*, *L. purvalus*, *H. bonvouloiri*, *O. cylindricus*, *O. cardoni*, *S. rifipes* and *C. marginipes* were found to be positively correlated with pH of the water revealing their preference for alkaline water. Similar observation has also been made by Jenila and Nair<sup>[27]</sup>. Two coleopteran species, *O. cardoni* and *S. rufipes* exhibited a significant positive correlation with DO while *P.*

*flavescens* (Odonata) had a significant negative correlation with DO. Pahari *et al.*<sup>[13]</sup> observed a significant negative correlation of two odonate species viz., *Anax imperator* and *Aeshna fabricius* with DO. Prommi and Payakka<sup>[28]</sup> reported that DO negatively affected taxa richness. While Barman and Gupta<sup>[1]</sup> observed significant positive correlation between species richness of aquatic insects and DO. Some species viz., *L. ruber*, *R. sordidula*, *M. scutellaris*, *D. annulatum*, *D. rusticus*, *P. liturata*, *C. luctuosus*, *L. purvulus*, *H. bonvouloiri*, *O. cylindricus*, *O. cardoni*, *S. rufipes*, *A. guttatus*, *C. marginipes*, *C. plumosus* showed a significant positive correlation with BOD. These are tolerant species and can survive in organically polluted water. *C. plumosus* is one such species. Several species like *R. sordidula*, *M. scutellaris*, *D. annulatum*, *P. liturata*, *C. luctuosus*, *B. contaminata*, *C. marginipes* and *C. plumosus* showed a significant positive correlation with TA while *A. bridini* alone exhibited a significant negative correlation with TA. Significant positive correlation between arthropods and bicarbonate alkalinity was also explained by Sharma and Chowdhary<sup>[29]</sup>. Interestingly none of the species exhibited any correlation with CO<sub>2</sub> of water. Nasiruddin *et al.*<sup>[30]</sup>, on the contrary opined that free CO<sub>2</sub> concentration and aquatic insect abundance is inversely correlated.

**Table 2:** Correlation coefficient between species and abiotic factors

Sl. No.	Specimen	Temperature	pH	DO	CO <sub>2</sub>	BOD	TA
1	<i>Laccotrephes ruber</i>	.484*	.734**	0.164	0.280	.704**	0.252
2	<i>Ranatra filiformis</i>	0.158	-0.174	0.027	0.344	0.378	0.174
3	<i>Ranatra sordidula</i>	.405*	.518**	0.268	0.152	.779**	.512*
4	<i>Anisops breddini</i>	.656**	-0.147	-0.146	0.054	-0.103	-.529**
5	<i>Micronecta scutellaris</i>	.431*	.739**	0.277	0.073	.776**	.453*
6	<i>Gerris spinolae</i>	0.225	-0.208	0.203	-0.091	0.280	0.210

7	<i>Diplonychus annulatum</i>	0.333	.510*	0.183	0.261	.520**	.629**
8	<i>Diplonychus rusticus</i>	.532**	0.390	0.165	0.264	.728**	0.308
9	<i>Lethocerus indicus</i>	0.214	.438*	0.217	0.385	0.210	0.369
10	<i>Plea liturata</i>	0.354	.644**	0.332	0.156	.772**	.566**
11	<i>Canthydrus luctuosus</i>	.444*	.638**	0.146	0.025	.681**	.578**
12	<i>Laccophilus purvulus</i>	.531**	.630**	0.225	0.347	.678**	0.272
13	<i>Hydrovatus bonvouloiri</i>	0.383	.507*	0.170	0.245	.649**	0.270
14	<i>Orectochilus cylindricus</i>	.551**	.627**	0.357	0.320	.789**	0.305
15	<i>Orectochilus cardoni</i>	0.357	.543**	.416*	0.371	.655**	0.372
16	<i>Sternolophus rufipes</i>	0.247	.585**	.568**	0.292	.579**	0.193
17	<i>Cloeon bicolor</i>	-.909**	-0.279	0.017	-0.121	-0.242	0.141
18	<i>Cloeon bengalense</i>	-.808**	-0.138	0.149	-0.082	-0.225	0.280
19	<i>Anaciaeschna jaspidea</i>	0.397	-0.008	-0.235	0.274	0.292	0.063
20	<i>Anax guttatus</i>	0.316	0.278	0.269	0.310	.462*	0.009
21	<i>Brachythemis contaminata</i>	0.208	0.403	0.146	0.159	0.394	.759**
22	<i>Crocothemis servilia</i>	.426*	0.074	-0.241	0.264	-0.191	-0.243
23	<i>Pantala flavescens</i>	0.334	0.250	-.466*	0.400	0.288	0.272
24	<i>Copera marginipes</i>	0.143	.473*	0.029	0.212	.513*	.779**
25	<i>Chironomus plumosus</i>	0.346	0.351	0.401	0.280	.651**	.447*

\*. Correlation is significant at the 0.05 level; \*\*. Correlation is significant at the 0.01 level.

Macrophyte diversity in the waterbody makes it a heterogenic habitat providing increased ecological niches that permitted co-existence of different insect species in such a small pond. Physico-chemical parameters ultimately determine the hypervolume niche.

#### Acknowledgement

Authors are grateful to the Principal of Tamralipta Mahavidyalaya and HOD, PG Department of Zoology for providing laboratory facilities.

#### References

- Barman B, Gupta S. Aquatic insects as bio-indicator of water quality-A study on Bakuamari stream, Chakras hila Wildlife Sanctuary, Assam, North East India. *Journal of Entomology and Zoology Studies*,2015;3(3):178–186.
- Subramanian KA, Sivaramakrishnan KG. A quatic insects for bio monitoring fresh w ater ecosystems: A methodology manual, 2007, Trust for Ecology and Environment (ATREE), Bangalore, India.
- Saha N, Aditya G, Saha GK. Prey preferences of aquatic insects: Potential implications for the regulation of wetland mosquitoes. *Medical and Veterinary Entomology*,2014;28(1):1-9. <https://doi.org/10.1111/mve.12003>
- Pahari PR, Chakraborty D, Mandal B, Bhattacharya T. Biological control of mosquito larvae using naiad of ruddy marsh skimmer *Crocothemis servilia*. *Indian Journal of Entomology*,2018;80(4):1503-1505. <https://doi.org/10.5958/0974-8172.2018.00330.9>
- Jacobus LM, Macadam CR, Sartori M. Mayflies (Ephemeroptera) and their contributions to ecosystem services. *Insects*,2019;10(6):170. <https://doi.org/10.3390/insects10060170>
- Sharma RK, Agrawal N. Faunal diversity of aquatic insects in Surha Tal of District - Ballia (U.P.), India. *Journal of Applied and Natural Science*,2012;4(1):60-64. <https://doi.org/10.31018/jans.v4i1.223>
- Majumder J, Das RK, Majumder P, Ghosh D, Agarwala BK. Aquatic Insect Fauna and Diversity in Urban Fresh Water Lakes of Tripura, Northeast India. *Middle East Journal of Scientific Research*,2013;13(1):25-32. 10.5829/idosi.mejsr.2013.13.1.66123
- Barman A, Baruah BK. Aquatic insects of Kapla Beel, a flood plain wetland of Barpeta District of Assam, India. *The Clarion International Multidisciplinary Journal*,2013;2(2):27-31.
- Choudhury D, Gupta S. Aquatic insect community of Deepor beel (Ramsar site), Assam, India. *Journal of Entomology and Zoology Studies*,2015;3(1):182-192.
- Choudhary A, Ahi J. Biodiversity of Freshwater Insects: A Review. *The International Journal of Engineering and Science*,2015;4:25–31.
- Pahari PR, Dutta, TK, Bhattacharya T. Aquatic insects of Midnapore district-II (Coleoptera: Hydrophilidae). *Vidyasagar University Journal of Biological Science*,1999;5:17-20.
- Pahari PR, Dutta TK, Bhattacharya T. Aquatic insects of Midnapore district-I (Insecta, Coleoptera, Dytiscidae). *Vidyasagar University Journal of Biological Science*,1997;3:45-51.
- Pahari PR, Pusti P, Dutta TK, Mandal B, Bhattacharya T. Diversity and Community Structure of Aquatic Insects in a Fresh Water Lentic System of Purba Medinipur District, W.B., India. *Indian Journal of Biology*,2016;3(2):145–150. <https://doi.org/10.21088/ijb.2394.1391.3216.9>
- Pahari PR, Dey M, Mishra NP, Bhattacharya T. Biodiversity of aquatic insects Aquatic Insects (Hemiptera) diversity in floating and emergent macrophytes in Garhbeta, West Bengal (India). *Int J Env Tech Sci*,2018;6:228–232.
- Pahari PR, Jana GC, Mandal S, Maiti S, Bhattacharya T. A Study on the Impact of Brick Embankment on Aquatic Entomofauna. *Uttar Pradesh Journal of Zoology*,2021;42(19):59–68.
- Jana S, Pahari PR, Dutta TK, Bhattacharya T. Diversity and community structure of aquatic insects in a pond in Midnapore town, West Bengal, India. *Journal of Environmental Biology*,2009;30(2):283-287.
- Engelmann HD. Untersuchungen zur Erfassung Predozoogener Komponenten im Definierten, Okosystem. *Forschungsergebnisse Staatliche Museum Naturkunde, Gorlitz. Journal of Acta Hydrobiologica*,1973;23(4):349–361.

18. APHA: Standard method of the examination of water and waste water. 21th Edn. APHA, AWWA and WPCF publication, 2005, Washington DC, USA.
19. Acuña V, Wolf A, Uehlinger U, Tockner K. Temperature dependence of stream benthic respiration in an Alpine river network under global warming. *Freshwater Biology*,2008;53(10):2076-2088. <https://doi.org/10.1111/j.1365-2427.2008.02028.x>
20. Dallas HF, Ketley ZA. Upper thermal limits of aquatic macroinvertebrates: Comparing critical thermal maxima with 96-LT50 values. *Journal of Thermal Biology*,2011;36(6):322-327. <https://doi.org/10.1016/j.jtherbio.2011.06.001>
21. Rotvit L, Jacobsen D. Temperature increase and respiratory performance of macroinvertebrates with different tolerances to organic pollution. *Limnologia*,2013;43(6):510-515. <https://doi.org/10.1016/j.limno.2013.04.003>
22. Resh VH, Rosenberg DM. Recent trends in life-history research on benthic macroinvertebrates. *Journal of the North American Benthological Society*, 2010, 29(1). <https://doi.org/10.1899/08-082.1>
23. Filipe AF, Lawrence JE, Bonada N. Vulnerability of stream biota to climate change in mediterranean climate regions: A synthesis of ecological responses and conservation challenges. *Hydrobiologia*,2013;719:331-351. <https://doi.org/10.1007/s10750-012-1244-4>
24. Eady BR, Rivers Moore NA, Hill TR. Relationship between water temperature predictability and aquatic macroinvertebrate assemblages in two South African streams. *African Journal of Aquatic Science*,2013;38(2):163-174. <https://doi.org/10.2989/16085914.2012.763110>
25. Rivers Moore NA, Dallas HF, Morris C. Towards setting environmental water temperature guidelines: A South African example. *Journal of Environmental Management*,2013;128:380-392. <https://doi.org/10.1016/j.jenvman.2013.04.059>
26. Adu BW, Oyeniyi EA. Water quality parameters and aquatic insect diversity in Aahoo stream, southwestern Nigeria. *The Journal of Basic and Applied Zoology*,2019;80(15):1-9. <https://doi.org/10.1186/s41936-019-0085-3>
27. Jenila GJ, Nair CR. Biodiversity of Aquatic Insect Population in Two Permanent Ponds of Kanyakumari District. *International Journal of Fauna and Biological Studies*,2013;1(2):8–12.
28. Prommi TO, Payakka A. Aquatic insect biodiversity and water quality parameters of streams in Northern Thailand. *Sains Mal*,2015;44:707-717.
29. Sharma KK, Chowdhary S. Macroinvertebrate assemblage as biological indicators of pollution in a central Himalayan river, Tawi (J & K). *Int. J. Bio. Con*,2011;3:167-174.
30. Nasiruddin M, Azadi MA, Saha S. Abundance and diversity of stream insects with reference to water quality. *Bangladesh J. Environ. Sci*,2015;28:13-22.