



Evaluation of toxicity of biopesticides against okra moth, *Earias vittella* (Fabricius) (Noctuidae: Lepidoptera)

Pratibha, Rajendra Singh

Department of Zoology, Deendayal Upadhyay Gorakhpur University, Gorakhpur, Uttar Pradesh, India

Abstract

The toxicity of garlic bulb aqueous extract, neem leaf aqueous extract and NeemGold (neem oil) (Azadiractin 0.03% EC, 300 ppm) was evaluated against okra moth, *Earias vittella* (F.) (Noctuidae: Lepidoptera) under laboratory conditions. The LC₅₀ values were determined at different doses (20, 30, 50 and 70 mg) were used for the aqueous extract of garlic bulb and neem leaf, and NeemGold (neem oil) for 24 h to 96 h exposure periods. The significant time and dose dependent toxic effect of all biopesticides were observed against *E. vittella*. The result demonstrated that *E. vittella* is very sensitive to NeemGold (LC₅₀ = 39.31 mg and 22.45 mg for 24 h and 96 h treatment, respectively) followed by neem leaf aqueous extract (LC₅₀ = 42.03 mg and 28.02 mg for 24 h and 96 h treatment, respectively) and garlic bulb aqueous extract (LC₅₀ = 61.85 mg and 39.42 mg for 24 h and 96 h treatment, respectively). It evinced that the NeemGold is more effective than neem leaf aqueous extract and garlic bulb aqueous extract for reducing the number of *E. vittella* larvae. Thus, the botanical tools are also valuable for managing the population of agricultural pests and being ecofriendly it gives better results for the management of *Earias vittella*.

Keywords: *Abelmoschus esculentus*, *Earias vittella*, Biopesticides, LC₅₀

1. Introduction

Okra (*Abelmoschus esculentus* (L.) Moench) is the only vegetable crop of significance in the Malvaceae family and is very popular in the Indo-Pak subcontinent [1]. It is not only the important source of vitamins and minerals but also proteins and carbohydrates. According to The okra pods can be consumed in different forms as well as important as fresh fruits [2]. It has also medicinal value and provide relief from ulcers and hemorrhoids [3]. Like other food crops, okra crop is also attacked by several insect pests among which spiny bollworm, *Earias vittella* (F.) (Noctuidae: Lepidoptera) is most injurious [4, 5]. *E. vittella* cause direct damage to tender fruits. The larvae attack and damage growing points of the vegetative parts, feed mostly inside squares, flowers and fruits. *E. vittella* causes 8.4-73.2% infestation on fruits of okra depending on the season [6]. Since, the synthetic insecticides cause health problems in humans, animals and also create environmental hazards, their field use should be minimum [7, 8]. In recent decades, effective uses of plant extracts against several insect pests have been reported [9, 10, 11, 12, 13]. Neem (*Azadirachta indica* A. Juss., Meliaceae) and garlic (*Allium sativum* L. var. *sativum*, Amaryllidaceae) based biopesticides have already shown their potentiality [14, 15]. Neem based biopesticides such as has systemic activity and it is even active at low concentrations but degrades rapidly in the environment [16]. Among the isolated Neem constituents, limonoids (azadirachtin) have multiple functions, such as insect growth regulation, affects the reproductive and digestive process of insects in such a manner that the insect can no longer feed, breed or undergo metamorphosis [17]. More specifically, it disrupts moulting by inhibiting biosynthesis or metabolism of ecdysone, the juvenile moulting hormone [18].

Garlic consists of varieties of volatile sulphur-based compounds which are effective as insect repellent as well as insecticides. Its one component, diallyl disulfide has a strong odour and acts as a powerful insecticide [5]. Commercial preparations of garlic are certified as insecticides against different pests infesting a variety of crops [19, 20, 21]. Aqueous garlic extracts have been shown to inhibit egg hatchability of mosquitoes [22], to have toxic and antifeedant effects on stored product pests [23] and Lepidoptera [24]. Repellent and toxic effects upon Hemiptera have also been observed [25, 24]. In India, *E. vittella* is one of the very common and notorious pest of okra. Therefore, the present study was undertaken to find out the LC_{50s} of aqueous extract of garlic bulb and neem leaf, and NeemGold against *E. vittella* larvae.

2. Materials and Methods

The *E. vittella* were reared following the method of Bhardwaj [26] in the laboratory. Under laboratory condition, the stock of *E. vittella* was maintained by procuring infested okra fruits from the local fields. As per requirement, different lots of such infested okra fruits were kept in rectangular insectaries (15x15x15 cm) made of card board, the top and four sides were fitted with soft fine mesh plastic net for ventilation. Inside the infested fruits, the larvae steadily grew till pupation by consuming the developing seeds. The full grown larva (approx. 2.0 cm in size) after completing development inside the okra fruit makes its exit and selects a suitable spot viz., the wall of the insectary, or even the outer surface of the fruit itself to pupate in a tough silken cocoon. Under optimal conditions, the pupal period lasts for 5 to 9 days at the end of which new generation of adult moths emerges after sunset.

Preparation of biopesticides

a. Aqueous extract of garlic bulbs (AEG): Aqueous extract of garlic bulb was prepared by grinding 1 kg of garlic bulbs with 1 l of distilled water. The extract was squeezed through fine meshed rayon cloth and finally filtered through Whatman filter paper. The filtrate (w/v) was used for foliar application as test biopesticide on experimental plant.

b. Aqueous extract of neem leaves (AEN): Leaves of neem were locally collected. Its aqueous extract was prepared by grinding 1 kg of fresh leaves with 1 l of distilled water. The extract was squeezed through fine meshed rayon cloth and finally filtered through Whatman filter paper. The filtrate (w/v) was used as test biopesticide for foliar application on experimental plant.

c. NeemGold preparation (NG): NeemGold was purchased from local market. It contains Azadiractin A 0.03%, Neem oil 90.57%, Hydroxy EL 5.00%, Epichorohydrin 0.50% and Aromex 3.90%, and is manufactured by Foliage Crop Solution Private Limited, Chennai, 600006, India. For preparation of its test solution, 1 ml of NeemGold was dissolved in 100 ml of distilled water.

Experimental set up

Above mentioned 3 biopesticides were used for their toxicity test, viz. AEG, AEN and NG. For each set, 20 third instar larvae of *E. vittella* were kept in an insectary and were kept in the laboratory at around 25°C. For every treatment of biopesticides, doses used were 20, 30, 50 and 70 mg/cage/20

larvae. After 24, 48, 72 and 96 h of exposure periods, the larvae were counted to record mortality. The lethal dose (LC₅₀), slope value, t-ratio and heterogeneity were calculated by the POLO computer programme following Robertson *et al.*, (2007). In the control set up, instead of biopesticides, only distilled water was sprayed over the larvae. The experiment was repeated 6 times for each biopesticide.

3. Results

The results are demonstrated in Table 1-3. The time and dose dependent toxic effect were observed in all the treatment of AEG, AEN and NeemGold in all the exposure periods (Fig. 1). Table 1 displays that the LC₅₀ values of AEG was 61.85 mg after 24 h which decreased upto 39.42 mg after 96 h of exposure period. Table 2 shows that the LC₅₀ values of AEN was 42.03 mg after 24 h of exposure period which decreased upto 28.02 mg after 96 h. Similarly, there was higher decrease in LC₅₀ value from 39.31 mg (24 h of exposure period) to 22.45 mg (96 h of exposure period) of NeemGold (Table 3). Regression coefficient showed a significant ($P < 0.05$) negative regression between exposure periods and different values of LC₅₀. The values of heterogeneity were very low and t-ratios are significant that demonstrate that the decrease in LC₅₀ value due to longer exposure period are significant, i.e., the toxicity of these biopesticides is time dependent. Enhanced exposure period increases the larval mortality in *E. vittella*. The data also demonstrated that the toxicity of NeemGold was much higher followed by AEN and AEG.

Table 1: Toxicity of garlic bulb aqueous extract againts *Earias vittella*.

Treated period (hrs)	Effective dose (LC ₅₀ mg)	Confident limits (mg)		Slope value	t- ratio	Heterogeneity
		LCL	UCL			
24	61.85	48.25	106.18	2.44±0.77	3.16	0.076
48	59.07	46.66	94.83	2.52±0.77	3.26	0.099
72	50.98	39.72	79.73	2.23±0.73	3.05	0.192
96	39.42	30.25	52.36	2.34±0.72	3.25	0.135

Table 2: Toxicity of neem leaf aqueous extract againts *Earias vittella*.

Treated period (hrs)	Effective dose (LC ₅₀ mg)	Confident limits (mg)		Slope value	t- ratio	Heterogeneity
		LCL	UCL			
24	42.03	33.41	55.14	2.56±0.73	3.509	0.035
48	36.92	28.55	46.98	2.54±0.72	3.50	0.024
72	31.29	22.61	39.24	2.50±0.73	3.423	0.028
96	28.02	20.11	34.51	2.76±0.75	3.664	0.283

Table 3: Toxicity of neem gold againts *Earias vittella*.

Treated period (hrs)	Effective dose (LC ₅₀ mg)	Confident limits (mg)		Slope value	t- ratio	Heterogeneity
		LCL	UCL			
24	39.31	31.82	48.97	2.90±0.74	3.895	0.068
48	33.34	24.65	42.15	2.46±0.72	3.391	0.029
72	29.29	20.67	36.59	2.54±0.73	3.438	0.061
96	22.45	14.46	28.06	2.89±0.816	3.553	0.244

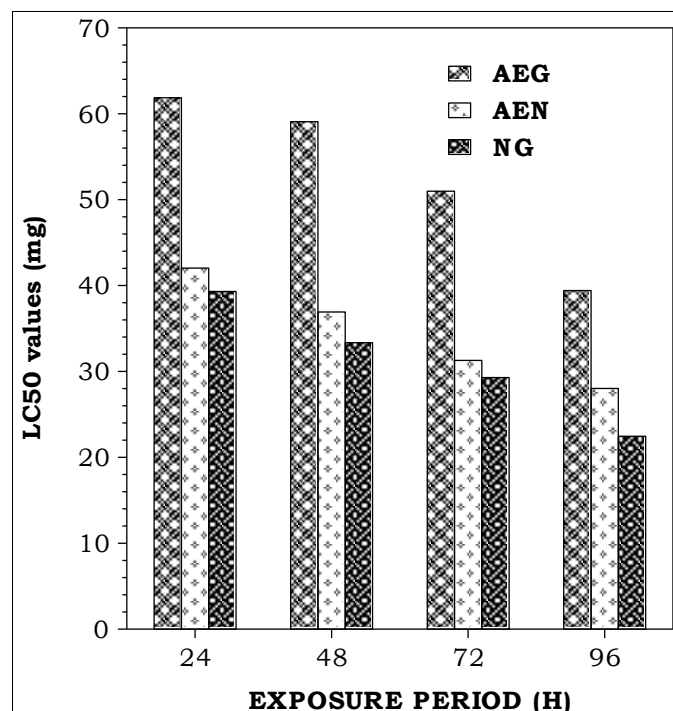


Fig 1: Relationship between LC₅₀ values of biopesticides tested and exposure periods.

4. Discussion

Biopesticides are certain types of pesticides derived from biological materials such as animals, plants, bacteria, etc. They are a set of tools and applications that help our farmers' transition away from highly toxic conventional chemical pesticides into an era of truly sustainable agriculture. India's rich biodiversity is an ace factor, always providing a wide source of biopesticides which can be effectively used in agriculture at a large scale. Also increasing health consciousness of Indian citizens has created a demand of organic food. This indicates huge scope for growth of biopesticides sector. Pesticidal property of garlic has earlier been reported against *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae) [27]. A 225.8 µl/l air of the garlic extract caused 83.3% larval mortality after 48 h. Even its very low dose (2.13 µl/cm²) caused more than 95% repellency. Two compounds extracted from garlic, (methyl allyl disulfide and diallyl trisulfide) have been observed toxic to other store grain beetles by reducing the egg hatchability and larval death [28, 29]. Bachrouch [11] also observed a dose dependent rate of egg mortality, higher dose of garlic extract increased the egg mortality. Garlic oil (2 %) protected the okra fruits from infestation of *E. vittella* when sprayed on the crop [30]. In other study of garlic essential oil was demonstrated to possess insecticidal activity against *Blattella germanica* (L.) (Blattodea: Blattellidae) [31]. Recently, Sohail [32] found reduced reproduction rate and high larval mortality of another okra moth *E. insulana* in Pakistan. Garlic oil caused substantial mortality and repellency in larva, pupa, and adult stages. The best results were obtained with concentrations of 16 and 32% in *Tenebrio molitor* (Coleoptera: Tenebrionidae) as reported for other stored grain pests according to the concentration of these products. [33, 34, 35]

A number of neem-based products have been observed to induce mortality of different kinds of insect pests on several vegetable and cereal crops [36, 37, 38, 12] and have been used in their management [39]. Its active ingredient, azadirachtin has been reported to reduce the fertility and fecundity of adults of green peach aphid *Myzus persicae* (Sulzer), lettuce aphid *Nasonovia ribisnigri* (Mosley) and strawberry aphid *Chaetosiphon fragaefolii* (Cockerell) in a linear concentration-dependent manner [40]. Neem-based insecticides have been found to deter feeding and thus influencing growth rate of beet armyworm (*Spodoptera exigua*) [41], *Trogoderma granarium* (Everts) (Coleoptera: Bostrichoidea) [42], *Cryptolestes pusillus* (Schon.) (Coleoptera: Polyphaga) [43]. Insecticidal influence of neem seed acetone extract was observed on the third instar larvae of *Corcyra cephalonica* (Staint.) (Lepidoptera: Pyralidae) and caused a depressive effect on the developmental stages of this moth. A dose level of 0.16% (a.i) v/w of this extract caused 100% larval mortality, which may be considered as extremely toxic to the pest [13]. Third instars larvae of the cabbage moth, *Mamestra brassicae* L. revealed that Neem EC inhibited larval development, greatly increased mortality, had antifeedant/deterrent effects and acted as a growth regulator for *Mamestra brassicae* (Lepidoptera: Noctuidae) larvae and pupae [44]. Experiments by Luik & Viidalepp [45] showed that when caterpillars of *Phytometra brassicae* (Lepidoptera: Noctuidae) were fed with cabbage treated with NeemAzal T/S, all caterpillars perished within 7 days. Active ingredient of neem i.e. azadirachtin inhibits the release of prothoracicotropic and allatotrophic hormones and hence affect the metamorphosis [46] and the growth [47]. Direct toxicity effect of neem based biopesticides (Neemazal, Neemarine, Bioneem, Neemgold, Nimbicidine and Achook) on *E. vittella* and *D. koenigii* was done by Kumar [48]. The result of toxicity effectiveness on the basis of the mortality ranged in descending order: Neemazal > Neemarine > Bioneem > Neemgold > Nimbicidine and Achook. Present study reveals that the biopesticides based on neem derivatives is more effective for the insect *E. vittella* than to garlic bulb extract.

5. Conclusion

It is concluded from the present experiments that the NeemGold (Azadirachtin 0.03% EC) is more potential in comparison to neem leaf extract and garlic bulb extract against *E. vittella*. Biopesticides are easily biodegradable, most effective and non hazardous to human and environment, leaves no residue and it is soft to insect predators and pollinators. Biopesticides may also offer solutions to concerns such as pest resistance to traditional chemical pesticides, public concern about side effects of pesticides on the surrounding environment and ultimately, on human health.

6. References

1. Kumar DS, Tony DE, Kumar AP, Kumar KA, Rao DBS, Nadendla R. A Review On: *Abelmoschus Esculentus* (Okra), International Research Journal Pharmaceutical Applied Science. 2013; 3(4):129-132.
2. Ndunguru J, Rajabu AC. Effect of okra mosaic virus disease on the above-ground morphological yield

- components of okra in Tanzania. *Scientia Horticulturae*. 2004; 99:225-235.
3. Adams CF. Nutritive value of American foods in common units, U.S. Department of Agriculture. *Agricultural Handbook*. 1975; 425:1-29.
 4. Krishnaiah K. Methodology for assessing crop losses due to pests of vegetables. In: Assessment of crop losses due to pests and diseases. Proc. of the workshop held at University of Bangalore, India, 1980, 259-267.
 5. Kaufman PB, Pennacchini L, McKenzie M, Hoyt JE. Good and bad uses of these compounds by humans. In: Kaufman, PB, Cseke LJ, Warber S, Dukes JA, Briellmann HL. (eds). *Natural products from plants*. New York: CRC Press. 1999, 123-56.
 6. Kumar KK, Urs KCD. Population fluctuation of *Earias vittella* (Fab.) on okra in relation to abiotic factor. *Indian Journal Plant Protection*. 1988; 16(2):137-142.
 7. Alavanja MC. Pesticides use and exposure extensive worldwide. *Review of Environmental Health*. 2009; 24:303-309.
 8. Arora NK, Tewari S, Singh S, Lal N, Maheshwari DK. PGPR for protection of plant health under saline conditions. In: Maheshwari DK (ed) *Bacteria in agrobiolgy: stress management*. Springer, Berlin, 2012, 239-258.
 9. Schmutterer H. Side effects of neem (*Azadirachta indica*) products on insect pathogens and natural enemies of spider mites and insects. *Journal of Applied Entomology*. 1997; 121(9):121-128.
 10. Carpinella MC, Defago MT, Valladares G, Palacios SM. Antifeedant and insecticide properties of a limonoid from *Melia azedarach* (Meliaceae) with potential use for pest management. *Agricultural Food Chemistry*. 2003; 15(51):369-674.
 11. Bachrouch O, Jema MB, Chaieb J, Talou I, Marzouk T, Abderraba B. Insecticidal activity of *Pistacia lentiscus* essential oil on *Tribolium castaneum* as alternative to chemical control in storage. *Tunisian Journal of Plant Protection*. 2010; 5:63-70.
 12. Akbar MF, Haq MA, Parveen F, Yasmin N, Farhan M, Khan U. Comparative management of cabbage aphid (*Myzus persicae* (Sulzer) (Aphididae: Hemiptera) through bio- and synthetic insecticides. *Pakistan Entomology*. 2010; 32(1):12-17.
 13. Pathak CS, Tiwari SK. Insecticidal Action of Neem Seed (*Azadirachta indica* A. Juss) Acetone Extract Against the Life-Cycle Stages of Rice-Moth, *Corcyra cephalonica* Staint. (Lepidoptera: Pyralidae) *World Journal of Agricultural Sciences*. 2012; 8 (5):529-536.
 14. Dhar Royal Dawar H, Garg S, Basir SF, Talwar GP. Effect of volatiles from neem and other natural products on gonotrophic cycle and oviposition of *Anopheles stephensi* and *An. Culicifacies* (Diptera: Culicidae). *Journal of Medical Entomology*. 1996; 33:195-201.
 15. Birrenkott GP, Brockenfelt GE, Greer JA, Owens MD. Topical application of garlic reduces northern fowl mite infestation in laying hens. *Poultry Science*. 2000; 79:1575-1577.
 16. Jacobson M. Focus on phytochemical insecticides. The neem tree. CRC Inc. Boca Raton, Florida, VSA. 1988; 1:178.
 17. Elahi KM. Social forestry, exotic trees and monga. The Daily Star Published 6 Sept 2008. URL: <http://www.thedailystar.net/story.php?nid=53438> (accessed on 6.
 18. Ware G, Whitacre D. An Introduction to Insecticides. In: Radcliffe E, Hutchison W, Cancelado R, Radcliffe's IPM World Texbook. URL: <http://ipmworld.umn.edu>. St. Paul (MN): University of Minnesota, 2004.
 19. Gupta R, Sharma NK. A study of the nematicidal activity of allacin: an active principle in garlic, *Allium sativum* L., against root-knot nematode, *Meloidogyne incognita*. *International Journal of Pest Management*. 1993; 39:390-2.
 20. Amonkar SV, Reeves EL. Mosquito control with active principle of garlic, *Allium sativum*. *Journal of Economical Entomology*. 1970; 63:1172-5.
 21. Gamboa-Leon R, Paraguai-de-Souza E, Borja-Cabrera GP, Santos FN, Myashiro LM, Pinheiro RO, Dumonteil E, Palatanik-de-Sousa CB. Immunotherapy against visceral leishmaniasis with the nucleoside hydrolase-DNA vaccine of *Leishmania donovani*. *Vaccine*. 2006; 24:4863-73.
 22. Jarial MS. Toxic effect of garlic extracts on the eggs of *Aedes aegypti* (Diptera: Culicidae): a scanning electron microscopic study. *Journal of Medical Entomology*. 2001; 38:446-450.
 23. Chiam WY, Huang Y, Chen SX, Ho SH. Toxic and antifeedant effects of allyl disulfide on *Tribolium castaneum* (Coleoptera: Tenebrionidae) and *Sitophilus zeamais* (Coleoptera: Curculionidae). *Journal of Economic Entomology*. 1999; 92:239-245.
 24. Gurusubramanian G, Krishna SS. The effects of exposing eggs of four cotton insect pests to volatiles of *Allium sativum* (Liliaceae). *Bulletin of Entomological Research*. 1996; 86:29-31.
 25. Flint HM, Parks NJ, Holmes JE, Jones JA, Higuera CM. Tests of garlic oil for the control of the silverleaf whitefly, *Bemisia argentifolia* Bellows and Perring (Homoptera: Aleyrodidae) in cotton. *Southwestern Entomologist*. 1995; 20:137-150.
 26. Bhardwaj AK, Ansari BA. Toxicity of Neem Based Pesticides (Nimbecidine and Neemazal) On the Cotton Pest, *Earias vittella*. *Scholars Academic Journal of Biosciences*. 2014; 2(11):768-772.
 27. Mobki M, Safavi SA, Safaralizadeh MH, Panahi O. Toxicity and repellency of garlic (*Allium sativum* L.) extract grown in Iran against *Tribolium castaneum* (Herbst) larvae and adults. *Phytopathology and plant protection*. 2014; 47(1):59-58.
 28. Huang Y, Chen SX, Ho SH. Bioactivities of methyl allyl disulfide and diallyl trisulfide from essential oil of garlic to two species of stored-product pests, *Sitophilus zeamais* (Coleoptera: Curculionidae) and *Tribolium castaneum* (Coleoptera: Tenebrionidae). *Journal Economical Entomology*. 2000; 93:537-543.
 29. Mona F, Aziz AE, Sayed YAE. Toxicity and biochemical efficacy of six essential oils against *Tribolium confusum* (Coleoptera: Tenebrionidae). *Egypt Academic Journal of Biological Sciences*. 2009; 2(2):1-11.

30. Sardana HR, Kumar NKK. Effectiveness of plant oils against leaf hopper and fruit and shoot borer on okra. *Indian Journal of Entomology*. 1989; 15(2):167-171.
31. Tunaz H, Er MK, Isikber AA. Fumigant toxicity of plant essential oils and selected monoterpenoid components against the adult German cockroach, *Blattella germanica* (L.) (Dictyoptera: Blattellidae). *Turkish Journal of Agriculture and Forestry*. 2009; 33:211-217.
32. Sohail K, Jan S, Usman A, Shah SF, Usman M, *et al.* Evaluation of some botanical and chemical insecticides against the insect pests of okra. *Journal of Entomology and Zoology Studies* 2015; 3 (2):20-24.
33. Isman, MB. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annual Review of Entomology*. 2006; 51:45-66.
34. De Menezes CWG, Tavares WS, De Souza EG, Soares MA, Serrão JE, Zanuncio JC. Effects of crude extract fractions of *Adenocalymma nodosum* (Bignoniaceae) on duration of pupa stage emergence of *Tenebrio molitor* (Coleoptera: Tenebrionidae) and phytotoxicity on vegetable crops. *Allelopathy Journal*. 2014; 33:141-150.
35. Mochiah M, Banful Fening K, *et al.* Botanicals for the management of insect pests in organic vegetable production. *Journal of Entomology and Nematology*. 2011; 3(6):85-97.
36. Lowery DT, Isman MB, Brard NL. Laboratory and field evaluation of neem for the control of aphids (Homoptera: Aphididae). *Journal Economic Entomology*. 1993; 86:864-870.
37. Coventry E, Allan EJ. Microbiological and chemical analysis of neem (*Azadirachta indica*) extracts: new data on antimicrobial activity. *Phytoparasitica*. 2001; 29(5):441-450.
38. Cutler CG, Ramanaidu K, Astatkie T, Isman MB. Green peach aphid, *Myzus persicae* (Homoptera: Aphididae), reproduction during exposure to sublethal concentrations of imidacloprid and azadirachtin. *Pest Management Science*. 2009; 65(2):205-209.
39. Yoshida H, Toscano NC. Comparative effects of selected natural insecticides on *Heliothis virescens* (Lepidoptera: Noctuidae) larvae. *Journal of Economic Entomology*. 1994; 87:305-310.
40. Lowery DT, Isman MB. Inhibition of aphid (Homoptera: Aleyrodidae) reproduction by neem seed oil and azadirachtin. *Journal of Economic Entomology*. 1996; 89: 602-607.
41. Greenberg SM, Showler AT, Liu TX. Effects of neem-based insecticides on beet armyworm (Lepidoptera: Noctuidae). *Insect Science*. 2006; 12:17-23.
42. Kapoor JS, Singh R. Evaluation of some plant products against *Trogoderma granarium* evverts in stored maize and their effects on nutritional composition and organoleptic characteristics of kernels. *Journal of Agricultural Chemistry*. 1993; 41:1644-1648.
43. Rhaman MM, Islam W, Sarker PK. Effect of azadirachtin on larvae and adults of *Cryptolestes pesillus* (Schon.) (Coleoptera: Cucujidae). *Pakistan Entomologist*. 2005; 27(1).
44. Metspalu L, Jõgar K, Ploomi A, Hiiesaar K, Kivimägi I, Luik A. Effects of biopesticide Neem EC on the *Mamestra brassicae* L. (Lepidoptera, Noctuidae) *Agronomy Research*. 2010; 2:465-470.
45. Luik A, Viidalepp V. On different modes of action of NeemAzal T/S on Cabbage butterfly. In Metspalu, L. & Mitt, S (eds): *Practice Oriented Results on the Use of Plant Extracts and Pheromones in Pest Control*, Tartu, 2001; 68-72.
46. Schmutterer H, Rembold H. *Neem tree source of unique natural products for Integrated pest management, medicine industry and other purposes* (ed. Schmutter, H.), VCH, Weinheim, Germany, 1995.
47. Banken JAO, Stark J. Stage and age influence on the susceptibility of *Coccinella septempunctata* after direct exposure to Neemex, a neem insecticide. *Journal of Economic Entomology*. 1997; 90: 1102-1105.
48. Kumar A. Toxic Effect of Neem Based Formulation against *Earias vitella* and *Dysdercus koenigii* Fabr. *International Journal of Science and Research*. 2013; 2(8).